

Autoimmune Rheumatic and Related Diseases

Laboratory Support for Classification and Diagnosis

Table of Contents

Clinical Background	1
Individuals Suitable for Testing	1
Test Availability	1
Test Selection and Interpretation.....	1
Gout and Pseudogout.....	1
Juvenile Idiopathic Arthritis.....	4
Mixed Connective Tissue Disease	4
Polymyositis and Dermatomyositis.....	4
Rheumatoid Arthritis.....	7
Sarcoidosis.....	8
Sjögren Syndrome	8
Spondyloarthropathies	9
Systemic Lupus Erythematosus and Neuropsychiatric Lupus.....	10
Systemic Sclerosis.....	10
Systemic Vasculitis.....	11
Appendix	12
Antinuclear Antibody Testing.....	12
Laboratory Tests for Classification and Diagnosis of Autoimmune Rheumatic and Related Diseases	13
References	17

CLINICAL BACKGROUND

Autoimmune rheumatic diseases (ARDs) are diseases in which the immune system attacks the joints and certain systems. The cause of many of these diseases is unknown. ARDs are sometimes difficult to distinguish owing to overlapping signs and symptoms: joint pain, diminished joint mobility, rash, fever, malaise, fatigue, and weight loss. Laboratory testing may be useful for the differential diagnosis and classification.

This Clinical Focus provides background on the available laboratory tests and their use in diagnosis and classification of the following autoimmune rheumatic and related diseases: gout and pseudogout, juvenile idiopathic arthritis (JIA), mixed connective tissue disease (MCTD), polymyositis and dermatomyositis (PM/DM), rheumatoid arthritis (RA), sarcoidosis, Sjögren syndrome, spondyloarthropathies (SpA), systemic lupus erythematosus (SLE) and neuropsychiatric lupus, systemic sclerosis (SSc), and systemic vasculitis. It does not cover laboratory testing as it relates to prognosis or treatment. It also does not comprehensively cover nonrheumatic autoimmune diseases (ie, Crohn disease, ulcerative colitis, autoimmune hepatitis) or nonautoimmune rheumatic diseases (ie, osteoarthritis), though some conditions that fit in these categories are covered (eg, gout, PM).

INDIVIDUALS SUITABLE FOR TESTING

Individuals who have signs and symptoms consistent with 1 or more ARD (**Table 1**).

TEST AVAILABILITY

Quest Diagnostics offers many tests and panels that may be useful for classifying or diagnosing ARDs (**Appendix Table**).

TEST SELECTION AND INTERPRETATION Gout and Pseudogout

The American College of Rheumatology (ACR)/European League Against Rheumatism (EULAR) classification criteria for gout (**Table 2**) include laboratory testing for monosodium urate (MSU) crystals and serum urate. However, a patient should only be tested if he or she has had at least 1 episode of pain, swelling, or tenderness in peripheral joints. If the patient has had an episode, the presence of MSU crystals in symptomatic joints indicates gout. If MSU crystals are absent, other criteria, including serum urate levels, are needed for a diagnosis. A high titer of serum urate is consistent with gout if other clinical or imaging criteria are met.¹

Pseudogout, also known as calcium pyrophosphate dihydrate (CPPD) crystal deposition disease, is definitively diagnosed by detection of CPPD crystals in synovial fluid or biopsy; diagnosis depends upon exclusion of other causes of arthritis.^{2,3}

Table 1. Common Signs and Symptoms of Autoimmune Rheumatic and Related Diseases^a

Sign or Symptom	Gout	JIA	MCTD	PM/DM	Pseudogout	RA	Sarcoidosis
Joint-/muscle-related							
Joint pain, stiffness, or inflammation	X	X	X	X	X	X	X
Muscle weakness			X	X			
Myalgia							
Skin-/hair-related							
Alopecia							
Rash		X	X	X			X
Raynaud phenomenon			X	X		O	
Skin lesions		X		O ^b			
General							
Anorexia						X	X
Cough							X
Ear involvement	O ^c						
Eye involvement		O					X
Fatigue				X		X	X
Fever	X	X	X	X		X	X
GI involvement							
Malaise	X					X	X
Nasal symptoms							O
Nervous system involvement							O
Respiratory involvement		X					X
Weight loss				X		X	X
Other							
Adenopathy		X					
Anemia						O	
Dysphagia				X			
Swelling of hands			X	O			O

(Continued)

Table 1. Common Signs and Symptoms of Autoimmune Rheumatic and Related Diseases^a (Continued)

Sign or Symptom	SjS	SLE	SpA				SSc	Systemic Vasculitis		
			AS	ReA	PsA	EA		GPA	EGPA	MPA
Joint-/muscle-related										
Joint pain, stiffness, or inflammation	X	X	X	X	X	X	X	X	X	X
Muscle weakness		O								
Myalgia		O						X	X	X
Skin-/hair-related										
Alopecia	X	X								
Rash	X	X							X	X
Raynaud phenomenon	X	X					X			
Skin lesions		O		O	X			O		O
General										
Anorexia			X							
Cough								X	X	
Ear involvement								X ^c		
Eye involvement	X	X	X	X				X		X
Fatigue		X	X	X					X	X
Fever		X	X	X				X	X	X
GI involvement							X	X	X	X
Malaise		X							X	X
Nasal symptoms								X	X	
Nervous system involvement		X						X	X	X
Respiratory involvement								X	X	
Weight loss			X	X					X	X
Other										
Adenopathy	X	X								
Anemia		X	X							
Dysphagia	X						X			
Swelling of hands					X		X			

X indicates common; O indicates less common but not rare. AS, ankylosing spondylitis; EA, enteropathic (inflammatory bowel disease-associated) arthritis; EGPA, eosinophilic granulomatosis with polyangiitis; GPA, granulomatosis with polyangiitis; JIA, juvenile idiopathic arthritis; MCTD, mixed connective tissue disease; MPA, microscopic polyangiitis; PM/DM, polymyositis/dermatomyositis; RA, rheumatoid arthritis; PsA, psoriatic arthritis; ReA, reactive arthritis; SjS, Sjögren syndrome; SLE, systemic lupus erythematosus; SpA, spondyloarthropathies; SSc, systemic scleroderma.

^a This is not a complete list of signs and symptoms; some conditions have more signs and symptoms than could be presented here.

^b In dermatomyositis.

^c External ear in gout; middle ear in GPA.

Table 2. Gout Classification Criteria¹

Classify a patient as having gout if:	
1. Patient has had ≥1 episode of pain, swelling, or tenderness in peripheral joint or bursa <i>and</i> monosodium urate crystals are present in symptomatic joint or bursa or tophus or	
2. Patient has had ≥1 episode of pain, swelling, or tenderness in peripheral joint or bursa <i>and</i> sum of points for criteria below is ≥8	
Criteria	Score
Clinical	
3. Pattern of joint involvement in monoarticular or oligoarticular episode	
– Involving ankle or midfoot (without involvement of first metatarsophalangeal joint)	1
– Involving first metatarsophalangeal joint	2
4. Characteristics of symptomatic episode	
– Erythema on affected joint	
– Touch or pressure on affected joint unbearable	1 per characteristic
– Inability to use affected joint	
5. Time course of typical episodes ^a	
– 1 typical episode	1
– Recurrent typical episodes	2
6. Presence of tophus ^b	4
Laboratory	
1. Serum uric acid: <4 mg/dL	-4
2. Serum uric acid: ≥4 to <6 mg/dL	0
3. Serum uric acid: 6 to 8 mg/dL	2
4. Serum uric acid: 8 to <10 mg/dL	3
5. Serum uric acid: ≥10 mg/dL	4
6. Monosodium urate negative in synovial fluid of symptomatic joint or bursa	-2
Imaging	
7. Ultrasound or dual-energy computed tomography (DECT) evidence of urate deposition in symptomatic joint or bursa	4
8. Radiographic evidence of gout-related joint damage (ie, erosion)	4

^a Typical episode is defined by ≥2 of the following, regardless of anti-inflammatory treatment: 1) maximal pain occurs in <24 hours; 2) symptoms resolve in ≤14 days; 3) symptoms completely resolve between symptomatic episodes.

^b Clinical evidence of tophus includes draining or chalk-like nodule under transparent skin, usually located in joints, ears, olecranon bursae, finger pads, or tendons.

Juvenile Idiopathic Arthritis

The International League of Associations for Rheumatology (ILAR) classification criteria define JIA as arthritis that begins before 16 years of age, persists for ≥6 weeks, and has unknown etiology.⁴ Although diagnosis of JIA is primarily clinical, ILAR recommends laboratory testing to distinguish between the forms of JIA (**Table 3**). For example, human leukocyte antigen (HLA)-B27 antigen testing and rheumatoid factor (RF) testing can help distinguish forms of JIA. A positive HLA-B27 antigen test result is consistent with enthesitis-related arthritis. A positive RF test result differentiates RF-positive and RF-negative forms of polyarthritis. Both of these test results also serve as exclusion criteria for other forms of JIA.

Mixed Connective Tissue Disease

Patients with MCTD can present with a wide range of signs and symptoms, most of which overlap with other ARDs. Four sets of MCTD classification criteria exist: Sharp, Alarcón-Segovia, Kasukawa, and Kahn.⁵ The different sets require a variety of clinical and serological criteria be met, but all 4 require either a positive result or a high titer for RNP antibody. For example, the Kasukawa criteria require a positive anti-RNP test result, whereas the Alarcón-Segovia criteria require a high RNP antibody titer (**Table 4**). The other sets of criteria factor in different laboratory test results, including a negative result for Smith antibody (Sm) and high titers of RNP or extractable nuclear antigen antibodies.

The first indication of MCTD is often a high antinuclear antibody (ANA) titer, which occurs in 94% to 97% of MCTD patients (see Appendix for more information about ANA testing).⁶ This test result should be followed by testing for antibodies to RNP, Sm, SS-A, SS-B, histone, and dsDNA.⁷ Over 90% of MCTD patients are positive for antibodies to RNP, while the other antibodies occur less frequently (<20% of patients).^{6,8} Antibodies to dsDNA, Sm, and SS-A can be seen transiently in MCTD, but consistent presence of these antibodies may indicate SLE.⁷

Polymyositis and Dermatomyositis

PM and DM are subgroups of idiopathic inflammatory myopathies (IIMs), which can be identified using the EULAR/ACR classification criteria for IIMs.⁹ These criteria include laboratory testing, including testing for Jo-1 antibody, creatine kinase, lactate dehydrogenase, aspartate aminotransferase, and alanine aminotransferase. A web-based calculator (<http://www.imm.ki.se/biostatistics/calculators/iim/>) that incorporates both clinical and laboratory criteria is available to estimate the probability that a patient has IIM. The

Table 3. Juvenile Idiopathic Arthritis Classification Criteria⁴

General		
1. Arthritis begins before 16 years of age 2. Arthritis persists ≥ 6 weeks 3. Other potential causes of arthritis are excluded		
JIA Form	Inclusion Criteria	Exclusion Criteria
Systemic arthritis	1. Arthritis in ≥ 1 joint <i>and</i> 2. Fever for ≥ 2 weeks that is daily for ≥ 3 days <i>and</i> 3. ≥ 1 of the following: 1. Evanescent erythematous rash 2. Generalized adenopathy 3. Hepatomegaly, splenomegaly, or both 4. Serositis	See footnotes a, b, c, and d.
Oligoarthritis	<i>Persistent form:</i> Arthritis in 1 to 4 joints during first 6 months of disease <i>and</i> in ≤ 4 joints during disease course <i>Extended form:</i> Arthritis in 1 to 4 joints during first 6 months of disease <i>and</i> in >4 joints after first 6 months	See footnotes a, b, c, d, and e.
Polyarthritis (RF-negative)	Arthritis in ≥ 5 joints during first 6 months of disease <i>and</i> negative results in RF test	See footnotes a, b, c, d, and e.
Polyarthritis (RF-positive)	Arthritis in ≥ 5 joints during first 6 months of disease <i>and</i> positive results in ≥ 2 RF tests run ≥ 3 months apart during first 6 months	See footnotes a, b, c, and e.
Psoriatic arthritis	1. Arthritis and psoriasis <i>or</i> 2. Arthritis and ≥ 2 of the following: 1. Dactylitis 2. Nail pitting or onycholysis 3. First-degree relative with psoriasis	See footnotes b, c, d, and e.
Enthesitis-related arthritis	1. Arthritis and enthesitis <i>or</i> 2. Arthritis or enthesitis <i>and</i> ≥ 2 of the following: 1. Sacroiliac joint tenderness and/or inflammatory lumbosacral pain 2. Positive for HLA-B27 antigen 3. Arthritis onset in male >6 years of age 4. Acute anterior uveitis 5. First-degree relative with ankylosing spondylitis, enthesitis-related arthritis, sacroiliitis with IBD, reactive arthritis, <i>or</i> acute anterior uveitis	See footnotes a, d, and e.
Undifferentiated arthritis	Arthritis that fulfills criteria for none of the above or ≥ 2 of the above types	NA

IBD, inflammatory bowel disease; JIA, juvenile idiopathic arthritis; NA, not applicable; RF, rheumatoid factor.

^a Psoriasis in patient or first-degree relative.

^b Arthritis beginning after 6th birthday in HLA-B27-positive male.

^c First-degree relative with ankylosing spondylitis, enthesitis-related arthritis, sacroiliitis with IBD, reactive arthritis syndrome, or acute anterior uveitis.

^d Positive results in ≥ 2 RF tests run ≥ 3 months apart.

^e Presence of systemic JIA.

Table 4. Mixed Connective Tissue Disease Diagnostic Criteria^{5,a}

Kasukawa Criteria	Alarcón-Segovia Criteria
<p>Diagnose MCTD if:</p> <ol style="list-style-type: none"> 1. RNP antibody test is positive <i>and</i> 2. ≥1 common symptom is present <i>and</i> 3. ≥1 mixed symptom in ≥2 disease categories <p>Common symptoms</p> <ol style="list-style-type: none"> 1. Raynaud phenomenon 2. Swollen fingers or hands <p>Mixed symptoms</p> <ol style="list-style-type: none"> 1. SLE-like symptoms (polyarthritits, lymphadenopathy, facial erythema, pericarditis or pleuritis, leukothrombocytopenia) 2. SSc-like findings (sclerodactyly, pulmonary fibrosis, restrictive changes of lung, reduced diffusion capacity, hypomotility or dilation of esophagus) 3. PM-like findings (muscle weakness, elevated serum levels of muscle enzymes [creatinine phosphokinase], myogenic pattern on electromyogram) 	<p>Diagnose MCTD if:</p> <ol style="list-style-type: none"> 1. RNP antibody titer >1:1,600 and 2. ≥3 clinical criteria present, including synovitis or myositis <p>Clinical criteria</p> <ol style="list-style-type: none"> 1. Edema in hands 2. Synovitis 3. Myositis 4. Raynaud phenomenon 5. 5Acrosclerosis

MCTD, mixed connective tissue disease; SLE, systemic lupus erythematosus; SSc, systemic sclerosis; PM, polymyositis.

^a Two of 4 existing MCTD criteria are shown here; they were selected because of their higher reported sensitivity and specificity (though the Kahn criteria perform similarly to Alarcón-Segovia criteria).^{5,6}

calculator also indicates type: PM, DM, amyopathic myositis, inclusion body myositis, and juvenile dermatomyositis.

Criteria relevant to subclassifying PM and DM are summarized in **Table 5**.

The classical myositis-specific antibodies include Jo-1, EJ, OJ, PL-7, and PL-12 synthetase antibodies, as well as Mi-2 and anti-signal recognition particle (SRP) antibodies. Of these, only Jo-1 antibody is currently included in the EULAR/ACR criteria.⁹ Others are anticipated to be included in future updates.⁹ Together, these antibodies are found in about 50% of patients with PM/DM and are often mutually exclusive.¹⁰

Jo-1 antibody is observed in 21% of patients with PM and in 11% of those with DM.¹⁰ It is 100% specific for PM/DM.^{11,12} The remaining classical myositis-specific antibodies occur less frequently than Jo-1¹⁰ but are also highly specific (97% to 100%) for PM/DM.^{11,13,14} Accordingly, a positive test result for any of the classical myositis-specific antibodies is highly suggestive of PM or DM (with rash). A negative test result does not rule out either PM or DM, however, as classical

myositis-specific antibodies are not detected in 50% of patients with these IIMs.

“Antisynthetase syndrome” is commonly observed in PM and DM patients. Arthralgia and interstitial lung disease are the

Table 5. PM and DM Classification Criteria^a

Classify patient as having PM (rash ^a absent) or DM (rash ^a present) if
<ul style="list-style-type: none"> • No other cause is present • Sum of EULAR/ACR classification points is ≥5.5 (≥6.7 with biopsy) • Age of onset is ≥18 years • Muscle weakness^b is present

DM, dermatomyositis; PM, polymyositis.

^a Heliotrope rash, Gottron sign, or Gottron papules.

^b Objective symmetric weakness, usually progressive of the upper or lower extremities; neck flexors weaker than extensors; or proximal leg muscles weaker than distal.

most prevalent extramuscular symptoms in patients with the syndrome.¹⁰ These patients typically have a moderate response to standard immunosuppressive therapy.¹⁵ Depending on the type of synthetase antibody, symptoms vary. A meta-analysis of 3,487 patients (27 studies) indicated that¹⁰

- Jo-1 antibody is associated with a significantly higher risk of myositis, arthralgia, and mechanic’s hand compared with non-Jo-1 antibodies.
- Non-Jo-1 antibodies (eg, EJ, OJ, PL-7, PL-12) are associated with a significantly higher risk of fever and interstitial lung disease compared with Jo-1 antibody.
- The risk of Raynaud phenomenon does not differ significantly between patients with Jo-1 antibodies and those with non-Jo-1 antibodies.

Rheumatoid Arthritis Rheumatoid Factor and Cyclic Citrullinated Peptide

The ACR/EULAR classification criteria for RA (**Table 6**) include testing for autoantibodies to RF and cyclic citrullinated peptide (CCP).¹⁶ RF is a widely used laboratory marker of RA. The reported sensitivity of RF is 57% for early RA¹⁷ and ranges from 60% to 86% for established RA.^{18,19} Positive RF results are suggestive of RA, but the relatively low specificity (70%-85%) precludes a definitive diagnosis for either early or established disease.^{17,19-23} Negative RF results are consistent with conditions other than RA but do not rule out RA; 14% to 43% of patients with RA are seronegative.¹⁷⁻¹⁹

The sensitivity of CCP antibody is comparable to that of RF in early (59%)¹⁷ and established RA (64%-88%).^{18,21} Unlike RF, CCP antibody is highly specific (90%-98%) for early and established RA.^{17,20,22,24} Most side-by-side comparisons demonstrate that CCP antibody is at least as sensitive as and more specific than RF in various clinical situations.^{18-21,23,25} Thus, positive CCP antibody results are highly suggestive of RA^{17,20,22,24}; however, patients with other rheumatic diseases may also have elevated titers. Negative results are consistent with RA and other rheumatic diseases. They do not rule out a diagnosis of RA; 12% to 41% of patients with RA are CCP seronegative.^{17,21} In RF-positive patients with chronic HCV or other infections associated with polyarticular arthritis, a

Table 6. Rheumatoid Arthritis Classification Criteria¹⁶

Classify a patient as having RA if sum of points is ≥ 6 .	
Criteria	Points
Joint involvement	
1. 1 large joint	0
2. 2-10 large joints	1
3. 1-3 small joints, with or without large joint	2
4. 4-10 small joints, with or without large joint	3
5. >10 joints with ≥ 1 small joint	5
Symptom duration	
1. <6 weeks	0
2. ≥ 6 weeks	1
RF and CCP antibody	
1. Normal RF and CCP antibody	0
2. Low-positive RF or CCP antibody	2
3. High-positive RF or CCP antibody	3
CRP and ESR	
1. Normal CRP and ESR	0
2. Elevated CRP or ESR	1

CCP, cyclic citrullinated peptide; CRP, C-reactive protein; ESR, erythrocyte sedimentation rate; RF, rheumatoid factor.

positive CCP antibody result suggests a likely diagnosis of coexisting RA; HCV patients with cryoglobulinemia, but not RA, typically have negative CCP antibody results.²⁶

The combination of RF and CCP antibodies provides greater sensitivity than either assay alone^{18,19,23} and is commonly used in the diagnostic evaluation of suspected RA. The combination of a positive IgM RF *and* CCP antibody result is highly suggestive of RA (~90%-100%). However, this test result may be found in some patients with other rheumatic diseases such as SLE, sclerosis, and psoriatic arthritis. Patients with negative RF and positive CCP antibody results are also likely to have RA. Patients with positive RF and negative CCP results are less likely to have RA, but RA remains a possibility. Negative results on both assays indicate a low likelihood of RA but do not exclude the diagnosis. Between 28% and 44% of patients with early disease test negative for both RF and CCP antibodies,^{27,28} which has led to the search for other RA markers.

14-3-3η

The 14-3-3η protein is elevated in serum and synovial fluid during joint inflammation and is a relatively new RA marker.²⁹ Although it is not yet included in the ACR/EULAR classification criteria, 14-3-3η antibody test sensitivity (64%) was higher than that of RF (57%) or CCP antibody (59%) in early RA patients in a side-by-side comparison.¹⁷ In patients with established RA, the sensitivity of 14-3-3η (77%) was comparable to that of RF or CCP antibody and specificity (93%) fell between that of RF and CCP antibody.¹⁷ Thus, positive 14-3-3η results are suggestive of RA.¹⁷ However, negative results do not rule out RA; 23% to 36% of patients with RA are 14-3-3η seronegative.¹² In a patient with a personal or family history of psoriasis, nail changes, and back or heel pain along with peripheral joint polyarthritis, a positive 14-3-3η result is consistent with a diagnosis of psoriatic arthritis.

Adding 14-3-3η testing to RF and CCP antibody testing provides greater sensitivity for early RA: 78% with 14-3-3η versus 72% without 14-3-3η.¹⁷ In seronegative patients, 14-3-3η detects 21% of patients with early RA and 67% of patients with established RA.¹⁷ This increased sensitivity may translate into treatment earlier in the course of disease, which can minimize irreversible joint damage. In patients with suspected RA, a positive/elevated result of RF, CCP antibody, and/or 14-3-3η protein suggests an RA diagnosis. Negative/normal results for all 3 markers indicate that an RA diagnosis is less likely.

C-reactive Protein and Erythrocyte Sedimentation Rate

C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) measurements are also included in the ACR/EULAR classification criteria for RA (**Table 6**). Elevated levels are consistent with an RA diagnosis if other laboratory and clinical criteria are met (**Table 6**). In patients with RA, elevated levels of CRP or ESR indicate heightened disease activity. However, elevations may also be due to other inflammatory conditions. Normal CRP and ESR results indicate relatively low disease activity. In patients with discordant CRP and ESR results, CRP levels may be the more reliable marker of RA disease activity.³⁰

Sarcoidosis

According to the American Thoracic Society (ATS), diagnosis of sarcoidosis requires 3 criteria (**Table 7**).³¹ The identification of noncaseating granulomas via biopsy is a strong indication of sarcoidosis; however, other causes of granulomatous disease must be excluded.³¹ Because other diseases must be excluded, laboratory tests may indirectly help with diagnosis.

Table 7. Sarcoidosis Diagnostic Criteria³¹

Diagnose sarcoidosis if patient has all of the following criteria:

1. Clinical or radiological presentation^a
2. Presence of noncaseating granulomas
3. Absence of alternative diseases^b

^a Chest x-ray or CT can identify hilar or mediastinal lymphadenopathy, upper lobe disease, subpleural reticulonodular infiltrates, peribronchial thickening, or traction bronchiectasis of the upper lobe.³³

^b Sarcoidosis shares signs and symptoms with other conditions including tuberculosis, prescription drug use (eg, methotrexate), granulomatous lesions of unknown significance (GLUS) syndrome, lymphoma, idiopathic pulmonary fibrosis, berylliosis, and multiple other infectious diseases.^{31,32}

Laboratory tests are not diagnostic for sarcoidosis, because they lack specificity, but some tests can support diagnosis. The following lab results are consistent with sarcoidosis: leukopenia, anemia, thrombocytopenia, elevated urinary calcium:creatinine ratio, and elevated serum levels of calcium, blood urea nitrogen (BUN), liver enzymes, immunoglobulins, or angiotensin-converting enzyme (ACE).³² The ATS recommends including peripheral blood counts, urine analysis, and serum levels of calcium, liver enzymes, creatinine, and BUN as part of a work-up once a diagnosis is confirmed.³¹

Sjögren Syndrome

The ACR and EULAR updated classification criteria for Sjögren syndrome in 2016.³⁴ The updated criteria involve inclusion criteria, exclusion criteria, and a scoring system based on 5 items of different weights (**Table 8**). The 2 most heavily weighted items are 1) SS-A/Ro testing and 2) labial salivary gland biopsy, which is a relatively invasive procedure that may be unnecessary if the other results are consistent with Sjögren syndrome. An initial validation indicates that sensitivity of the 2016 criteria is 96% (95% CI 92%-98%) and specificity is 95% (95% CI 92%-97%).³⁴

Previous classification criteria from 2012 included testing for SS-B/La, RF, and ANA titer in addition to SS-A/Ro.³⁵ Those criteria indicated that testing for SS-A or SS-B antibody has a sensitivity of 84% and specificity of 92%, and that a positive RF result (sensitivity, 72%; specificity, 86%) in addition to an ANA titer ≥1:320 (sensitivity, 73%; specificity, 80%) was consistent with the Sjögren syndrome. These markers were not included in 2016 guidelines, because studies indicated the markers do not add sufficient value to SS-A/Ro testing.³⁴

Table 8. Sjögren Syndrome Classification Criteria³⁴

Classify as Sjögren Syndrome if	
Patient meets ≥ 1 of the inclusion criteria	
<ol style="list-style-type: none"> Ocular or oral dryness^a Suspicion of Sjögren syndrome based on EULAR Disease Activity Index questionnaire³⁶ 	
.....	
Patient does <i>not</i> meet any of the exclusion criteria ^b	
<ol style="list-style-type: none"> Radiation treatment of head and neck Active hepatitis C infection (confirmed by PCR) AIDS Sarcoidosis Amyloidosis Graft-versus-host disease IgG4-related disease 	
.....	
Patient scores ≥ 4 based on the weights of the following items (weights in parentheses)	
<ol style="list-style-type: none"> Labial salivary gland biopsy exhibits focal lymphocytic sialadenitis with focus score ≥ 1 focus/4 mm² (3 points) SS-A/Ro test is positive (3 points) Ocular staining score is ≥ 5 (or van Bijsterveld score ≥ 4) in ≥ 1 eye Schirmer's test ≤ 5 mm/5 minutes in ≥ 1 eye Unstimulated whole saliva flow rate ≤ 0.1 mL/min 	

EULAR, European League Against Rheumatism; PCR, polymerase chain reaction.

^a Defined as having or doing ≥ 1 of the following: dry eyes for >3 months; recurrent sensation of sand or gravel in eyes; use of tear substitutes ≥ 3 times per day; feeling of dry mouth every day for >3 months; often drink to help swallow food.

^b Clinical features of these conditions overlap with SS and interfere with criteria tests.

Spondyloarthropathies

The term “spondyloarthropathies” (SpA) encompasses a group of inflammatory rheumatic diseases that cause arthritis; these include ankylosing spondylitis, reactive arthritis, psoriatic arthritis, and inflammatory bowel disease-associated arthritis. SpA can be subdivided into axial SpA, which involves the spine and sacroiliac joints, and peripheral SpA, which involves peripheral arthritis, enthesitis, and dactylitis.^{37,38} Overlapping presentations involving both axial and peripheral joints may occur.

The Assessment of SpondyloArthritis International Society (ASAS) has released classification criteria for axial SpA and peripheral SpA.^{37,38} Laboratory testing for HLA-B27 can help identify individuals with SpA.^{37,38} The axial SpA criteria have radiographic (imaging) and clinical arms, both of which incorporate HLA-B27 testing (**Table 9**); these criteria (ie, fulfilling criteria of either arm) have a sensitivity of 83% and a specificity of 84%.³⁸ The peripheral SpA criteria, which also incorporate HLA-B27 testing, have a sensitivity of 78% and a specificity of 82%.³⁷

A positive HLA-B27 result is consistent with any type of SpA (ankylosing spondylitis, reactive arthritis, psoriatic arthritis, or inflammatory bowel disease-associated arthritis), acute anterior uveitis, or JIA. However, most people who are HLA-B27 positive do not develop an associated condition. Thus, diagnosis and classification should be based on multiple criteria.^{37,38} A negative HLA-B27 result does not rule out conditions associated with HLA-B27; the allele is not always present in patients with these conditions.

CRP levels can also assist with classification of axial SpA. Elevated levels are consistent with axial SpA in the presence of other criteria. A negative result does not rule out a classification of SpA since other criteria may be met.

Table 9. Spondyloarthropathy Classification Criteria

Axial SpA ³⁸		Peripheral SpA ³⁷
Patients with back pain for ≥ 3 months who are <45 years at onset and meet criteria in clinical or imaging arm		Patients with peripheral manifestations only
Clinical Arm	Imaging Arm	
HLA-B27 and ≥ 2 other SpA features from footnote a	Sacroiliitis on imaging <i>and</i> ≥ 1 SpA feature from footnote a	Arthritis, enthesitis, or dactylitis <i>and</i> ≥ 1 SpA feature from footnote b <i>or</i> ≥ 2 other SpA features from footnote c

^a HLA-B27, inflammatory back pain, arthritis, enthesitis, uveitis, dactylitis, psoriasis, Crohn disease or ulcerative colitis, response to NSAIDs, family history of SpA, elevated C-reactive protein levels.

^b HLA-B27, uveitis, psoriasis, Crohn disease or ulcerative colitis, preceding infection, sacroiliitis on imaging.

^c Arthritis, enthesitis, dactylitis, inflammatory back pain, family history of SpA.

Systemic Lupus Erythematosus and Neuropsychiatric Lupus

Classification of SLE according to the Systemic Lupus International Collaborating Clinics (SLICC) criteria requires a combination of clinical and immunologic evaluation (Table 10).³⁹ Laboratory testing can help assess some clinical criteria such as the presence of proteinuria, anemia,

Table 10. SLE Classification Criteria³⁹

Classify a patient as having SLE if:

1. 4 criteria are met, including ≥ 1 clinical and ≥ 1 immunologic criterion *or*
2. Biopsy-proven nephritis compatible with SLE *and* ANA *or* dsDNA antibodies are present

Clinical Criteria

1. Acute cutaneous lupus in the absence of dermatomyositis *or* subacute cutaneous lupus
2. Chronic cutaneous lupus
3. Oral ulcers^a *or* nasal ulcers^a
4. Nonscarring alopecia^a
5. Synovitis of ≥ 2 joints *or* tenderness of ≥ 2 joints and >30 minutes morning stiffness
6. Pleurisy (>1 day), pleural effusion, *or* pleural rub^a *or* pericardial pain (>1 day), pericardial effusion, pericardial rub, *or* pericarditis by ECG^a
7. Urine protein-to-creatinine ratio indicates 500 mg protein/24 hours *or* red blood cell casts
8. Seizures, psychosis, myelitis, mononeuritis multiplex,^a peripheral *or* cranial neuropathy,^a *or* acute confusional state^a
9. Hemolytic anemia
10. Leukopenia ($<4,000/\text{mm}^3$)^a *or* lymphopenia ($<1,000/\text{mm}^3$)^a
11. Thrombocytopenia ($<100,000/\text{mm}^3$)^a

Immunologic Criteria

1. ANA level above reference range
2. dsDNA antibody level above reference range (*or* $>$ twice reference range if tested by ELISA)
3. Sm antibody positive
4. Antiphospholipid antibody positive^b
5. Low C3, C4, *or* CH50
6. Direct Coombs test if hemolytic anemia is absent

^a If no other cause is present.

^b As determined by positive result for lupus anticoagulant; false-positive result for rapid plasma reagin; medium- to high-titer of cardiolipin antibody; *or* positive results for $\beta 2$ -glycoprotein I antibody.

leukopenia, lymphopenia, and thrombocytopenia. It can also help assess immunologic criteria such as the presence of antibodies to DNA, Sm, antiphospholipids, and ANA, as well as levels of complement proteins (C3, C4, and total).

When considered individually, most of the laboratory tests included in the SLICC classification criteria provide high specificity (86% to 99%) and low to medium sensitivity for SLE (7% to 59%). The exception is ANA testing, which has medium specificity (45%) and high sensitivity (96%)³⁹; a negative ANA test can help rule out SLE because ANA-negative SLE is rare.⁴⁰

Though not included in the SLICC classification criteria, chromatin antibodies have relatively high sensitivity (64% to 69%) and specificity (92% to 99%) for SLE,^{41,42} and may provide value when diagnosing SLE. RNP antibodies are also present in SLE patients, but are not specific to SLE; RNP antibodies are more useful for identifying MCTD.^{43,44}

Neuropsychiatric lupus has a broad range of symptoms in the central *or* peripheral nervous systems. Validated diagnostic criteria do not exist, but diagnosis usually involves a clinical evaluation that is supported by results from brain MRI, serology and cerebrospinal fluid (CSF) testing, and neuropsychiatric assessment; diagnosis also requires the exclusion of other potential causes of neuropsychiatric signs and symptoms.⁴⁰ Laboratory testing can indirectly help diagnose neuropsychiatric lupus by excluding other causes, including infectious diseases. In addition, compared to SLE patients without neuropsychiatric disease, patients with neuropsychiatric lupus often have higher serum levels of neuronal antibodies *or* antibodies to ribosomal P, cardiolipin, lupus anticoagulants, *or* phospholipids.⁴⁵

Systemic Sclerosis

The EULAR/ACR classification criteria for SSc (Table 11) comprise mainly clinical criteria, but also include laboratory testing for autoantibodies.⁴⁶ A positive result for centromere, Scl-70 (topoisomerase I), *or* RNA polymerase III antibodies is consistent with SSc but is not diagnostic. A positive ANA result suggests the need for testing for specific autoantibodies if clinical symptoms are consistent with SSc; 85% to 97% of patients with SSc are ANA-positive (see Appendix for more information about ANA testing).^{47,48}

The 2 most common types of SSc are diffuse cutaneous SSc (dcSSc) and limited cutaneous SSc (lcSSc), also called CREST (calcinosis, Raynaud phenomenon, esophageal dysmotility, sclerodactyly, telangiectasia) syndrome. The type of SSc can affect prognosis and treatment. Differentiation may

be possible based on location of skin fibrosis (proximal vs distal extremities) and clinical manifestations, but laboratory test results can support differentiation. Scl-70 antibody is found in approximately 40% of patients with dcSSc, whereas centromere antibody is found in up to 90% of patients with lcSSc.⁴⁹ When detected by indirect immunofluorescence, immunoprecipitation, or immunodiffusion, Scl-70 and centromere antibodies are almost always mutually exclusive in SSc patients: only 0.5% test positive for both.⁴⁹ Thus, a positive test result for Scl-70 antibody is consistent with dcSSc if clinical symptoms are present, and a positive test result for centromere antibody is consistent with lcSSc if clinical symptoms are present.

Systemic Vasculitis

The ACR has created classification criteria for 2 autoimmune systemic vasculitis disorders (**Table 12**): granulomatosis with polyangiitis (GPA, formerly Wegener granulomatosis)⁵⁰ and eosinophilic granulomatosis with polyangiitis (EGPA, formerly Churg-Strauss syndrome).⁵¹ Laboratory tests are included in both sets of criteria. If GPA is suspected, testing for microhematuria can assist with classification. If EGPA is suspected, testing for eosinophilia can assist with classification. Other routine laboratory test results can suggest systemic vasculitis. Anemia, leukocytosis, thrombocytosis, and elevated ESR and CRP levels are consistent with an acute phase response.⁵²

Diagnosis should be confirmed via biopsy of the affected tissue when possible; this is true for GPA, EGPA, and another autoimmune systemic vasculitis disorder, microscopic polyangiitis (MPA). Classification criteria for MPA are not published, but clinical and histological findings in patients with MPA include fibrinoid necrotizing vasculitis

of predominantly small vessels without immune deposits, focal segmental necrotizing glomerulonephritis, pulmonary capillaritis, and neutrophilic infiltration of the alveolar wall.⁵²

Table 11. Systemic Sclerosis Classification Criteria⁴⁶

Classify a patient as having systemic sclerosis if sum of points is ≥ 9 .	
Criteria	Points
Skin thickened on fingers of both hands, extending proximal to the metacarpophalangeal joints	9
Skin on fingers thickened (only count highest score)	
– Puffy fingers	2
– Sclerodactyly ^a	4
Lesions on fingertips (only count highest score)	
– Ulcers on tip of digits	2
– Pitting scars on fingertips	3
Telangiectasia	2
Abnormal nailfold capillaries	2
Pulmonary arterial hypertension and/or interstitial lung disease (max score is 2)	
– Pulmonary arterial hypertension	2
– Interstitial lung disease	2
Raynaud phenomenon	3
Presence of any of the following SSc-related autoantibodies ^b : centromere, Scl-70, or RNA polymerase III	3 ^b

^a Distal to metacarpophalangeal joints but proximal to proximal interphalangeal joints.

^b 3 points for 1 or more of the antibodies; maximum score is 3.

Table 12. Granulomatosis with Polyangiitis and Eosinophilic Granulomatosis with Polyangiitis Classification Criteria

Classify patient as having granulomatosis with polyangiitis if ≥ 2 of the following are present ⁵⁰ :	Classify patient as having eosinophilic granulomatosis with polyangiitis if ≥ 4 of the following are present ⁵¹ :
1. Oral ulcers or bloody or purulent nasal discharge	1. Asthma
2. Nodules, fixed infiltrates, or cavities on chest radiograph	2. Eosinophilia $>10\%$
3. Microhematuria or red cell casts in urine sediment	3. Mono- or polyneuropathy
4. Granulomatous inflammation in wall of artery or peri- or extra-vascular area	4. Nonfixed pulmonary infiltrates
	5. Paranasal sinus abnormality
	6. Extravascular eosinophils

Differential diagnosis of GPA, EGPA, and MPA can be aided by testing for specific antineutrophil cytoplasmic antibodies (ANCA).⁵³ Each disorder is associated with predominance of a specific ANCA type.⁵⁴ The ANCA types are revealed by fluorescent patterns obtained in an indirect immunofluorescence ANCA screen. For example, the cytoplasmic pattern (C-ANCA) is very common in GPA, but not MPA or EGPA. The perinuclear pattern (P-ANCA), on the other hand, is rare in GPA, common in MPA, and moderately common in EGPA cases. The atypical P-ANCA pattern is rare in all 3 of these; it is usually associated with nonvasculitic conditions such as inflammatory bowel disease.⁵⁵ The sensitivity and specificity of these markers for the various disorders are summarized in (Table 13).⁵⁶⁻⁵⁸

Table 13. Diagnostic Accuracy of Antibodies for Systemic Vasculitis

Markers	% Sensitivity (Specificity)		
	GPA ^{58,59,a}	MPA ^{58,59,a}	EGPA ⁵⁶
ANCA	85 (93)	68 (87)	31
C-ANCA	81 (100)	3 (93)	5
C-ANCA+/PR3+	69 (100)	0	1
PR3+	77-81 (98-99)	9-12 (96-99)	
P-ANCA	4 (94)	65 (94)	21
P-ANCA+/MPO+	2 (99)	48 (100)	20
MPO+	5-9 (98-99)	71-88 (96-99)	

ANCA, antineutrophil cytoplasmic antibodies; EGPA, eosinophilic granulomatosis with polyangiitis; GPA, granulomatosis with polyangiitis; MPA, microscopic polyangiitis.

^a Sensitivity and specificity based on patients tested for ANCA in a rheumatology clinic.

An international consensus group recommends improving the diagnostic accuracy of the ANCA screen by combining it with immunoassays specific for myeloperoxidase (MPO) and proteinase-3 (PR3) antibodies (Table 13).⁵⁴ The P-ANCA pattern predominantly reflects MPO specificity. Similarly, the C-ANCA pattern typically reflects specificity to PR3; however, concordance between C-ANCA and PR3 antibody is not 100% because C-ANCA has multiple targets.

Alternatively, a more recent international consensus statement recommends that high quality MPO- and PR3-immunoassays be used as a primary screening method for GPA and MPA, based on the improved performance of these assays (Table 13).⁵⁹ Positive PR3 assay results are highly suggestive of GPA and positive MPO results are highly suggestive of MPA. The

group proposes that indirect immunofluorescence only be used in patients with negative MPO- and PR3-assay results and a high clinical suspicion of disease.⁵⁹

A positive ANCA screen supports a diagnosis of autoimmune-related systemic vasculitis in a symptomatic patient (Table 13). Positive results are also seen in inflammatory bowel disease (ulcerative colitis) and occasionally in other autoimmune diseases (SLE, RA, autoimmune hepatitis). Exposure to certain drugs (eg, propylthiouracil, hydralazine, methimazole) and infectious agents (eg, hepatitis C virus) can result in secondary vasculitis and an ANCA-positive screen result.^{60,61} A negative ANCA, MPO antibody, and/or PR3 antibody result does not rule out systemic vasculitis.

Owing to limitations in sensitivity and specificity, ANCA, MPO antibody, and PR3 antibody test results should be interpreted carefully in light of clinical and other laboratory data.

APPENDIX Antinuclear Antibody Testing

The laboratory work-up for patients with suspected rheumatic disease often begins with an antinuclear antibody (ANA) screen. The classic ANA testing approach uses HEp-2 human tissue culture cells in an immunofluorescence assay (IFA) to detect autoantibodies directed against antigens in the cell cytoplasm and nucleus. Because it is highly sensitive, this method is considered by the American College of Rheumatology (ACR) to be the current gold standard.⁴³

The high sensitivity of IFA stems from inclusion of a large number of antigens; however, these antigens are not very disease specific. Analyte-specific immunoassays are more disease specific owing to the use of antigens strongly associated with particular rheumatic diseases. Multiplex immunoassays can identify multiple autoantibodies simultaneously, but the number of antibodies detected is fewer than in the IFA; this difference results in lower sensitivity. Thus, the IFA and immunoassay methods complement each other for the initial evaluation of suspected rheumatic disorders. When ANA is positive by IFA, especially with high titer, testing for disease-specific antibodies can help with differential diagnosis.

Samples with an IFA titer <1:40 are considered negative for ANA antibodies; follow-up with more specific testing is not needed. Higher titers are generally associated with greater likelihood of rheumatic disease, but do not reflect disease activity. When results are positive, various fluorescent staining patterns are observed in the nucleus or the cytoplasm. These patterns can aid in the differential diagnosis of rheumatic disease and guide selection of further testing for specific autoantibodies.

Table. Laboratory Tests for Classification and Diagnosis of Autoimmune Rheumatic and Related Diseases^a

Test Code	Test Name	Clinical Use
Gout and Pseudogout		
4563	Crystals, Synovial Fluid	Diagnose gout and pseudogout
Juvenile Idiopathic Arthritis		
4420	C-Reactive Protein (CRP)	Diagnose axial spondyloarthritis and assess disease activity
528	HLA-B27 Antigen	Diagnose spondyloarthropathies
Mixed Connective Tissue Disease		
19875 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Mixed Connective Panel 1 Includes ANA screen (IFA) with a reflex to titer and pattern and RNP antibody.	Diagnose MCTD
90074 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Mixed Connective Panel 2 Includes ANA screen (IFA) with reflexes to titer and pattern; dsDNA, RNP, and Scl-70 antibodies.	Diagnose MCTD
19887	RNP Antibody	Diagnose SLE or MCTD
38567	Sm/RNP Antibody	Diagnose SLE or MCTD
Polymyositis and Dermatomyositis		
823	Alanine Aminotransferase (ALT)	Diagnose PM/DM
227	Aldolase	Diagnose PM/DM
38075	Antisynthetase Antibody Panel 1 Includes Jo-1 (test code 5810[X]), EJ (test code 90998), OJ (test code 90999), PL-7 (test code 90996), and PL-12 (test code 90997) antibodies.	Diagnose antisynthetase syndrome in a symptomatic patient
822	Aspartate Aminotransferase (AST)	Diagnose PM/DM
374	Creatine Kinase (CK), Total	Diagnose PM/DM
5810(X)	Jo-1 Antibody	Diagnose PM/DM
593	Lactate Dehydrogenase (LD)	Diagnose PM/DM
90995	Myositis AssessR TM Includes EJ (test code 90998), OJ (test code 90999), PL-7 (test code 90996), PL-12 (test code 90997), Mi-2 (test code 17172), Ku (test code 18855), and SRP (test code 16318) antibodies.	Diagnose PM/DM
10185(X)	Myositis AssessR TM plus Jo-1 Antibodies Includes Antisynthetase Antibody Panel 1 (test code 38075), Mi-2 (test code 17172), Ku (test code 18855), and SRP (test code 16318) antibodies.	Diagnose PM/DM (Jo-1 provides a more definitive diagnosis)
94777 ^c	Myositis Specific 11 Antibodies Panel Includes Jo-1, EJ, OJ, PL-7, PL-12, SRP, Mi-2 α , Mi-2 β , MDA-5, NXP-2, and TIF1- γ antibodies.	Diagnose PM/DM (Jo-1 provides a more definitive diagnosis) Diagnose NM Diagnose cancer-associated DM Diagnose juvenile DM Diagnose amyopathic DM
Rheumatoid Arthritis		
91455 ^c	14.3.3 eta Protein	Diagnose RA; more sensitive than either RF or CCP for early RA ¹⁷

(Continued)

Table. Laboratory Tests for Classification and Diagnosis of Autoimmune Rheumatic and Related Diseases^a (Continued)

Test Code	Test Name	Clinical Use
90071 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Rheumatoid Arthritis Panel 1 Includes ANA screen (IFA) with reflex to titer and pattern; cyclic citrullinated peptide (CCP) IgG; rheumatoid factor.	Diagnose RA
92813 ^{b,c}	ANA Screen, IFA, with Reflex to Titer and Pattern/ (Rheumatoid Arthritis Panel 2) Includes ANA screen (IFA) with reflex to titer and pattern, 14-3-3 eta protein; cyclic citrullinated peptide (CCP) IgG; rheumatoid factor.	Diagnose RA
4420	C-Reactive Protein (CRP)	Diagnose RA and assess disease activity
11173	Cyclic Citrullinated Peptide (CCP) Antibody (IgG)	Diagnose and determine prognosis of RA; more specific than RF
809	Sed Rate b Modified Westergren	Diagnose RA and assess disease activity
657	Mucin Clot, Synovial Fluid	Differential diagnosis of diseases of joints and joint fluid
91472 ^c	Rheumatoid Arthritis Diagnostic IdentRA [®] Panel 2 Includes 14-3-3 eta protein, cyclic citrullinated peptide (CCP) IgG, and rheumatoid factor.	Diagnose RA; provides additional diagnostic and prognostic value relative to each assay alone
92812 ^c	Rheumatoid Arthritis Diagnostic IdentRA [®] Panel 4 Includes 14-3-3 eta protein, cyclic citrullinated peptide (CCP) IgG, rheumatoid factor antibodies (IgA, IgG, IgM), and SS-A and SS-B antibodies.	Diagnose RA; may help differentiate RA from RA with secondary Sjögren syndrome
17669	Rheumatoid Arthritis Diagnostic Panel 1 Includes cyclic citrullinated peptide (CCP) IgG and rheumatoid factor.	Diagnose RA
19878	Rheumatoid Arthritis Diagnostic Panel 3 Includes cyclic citrullinated peptide (CCP) IgG; rheumatoid factor (IgA, IgG, and IgM); and SS-A and SS-B antibodies.	Diagnose RA; may help differentiate RA from RA with secondary Sjögren syndrome
4418	Rheumatoid Factor	Diagnose and determine prognosis of RA; detects primarily IgM RF
15682	Rheumatoid Factor (IgA)	Diagnose RA; provides added specificity when used in combination with other RF or CCP antibody assays; may help predict severity of disease course
19705	Rheumatoid Factor (IgA, IgG, IgM)	Diagnose RA; detecting all 3 isotypes improves specificity and predictive value
15683	Rheumatoid Factor (IgG)	Diagnose RA; provides added specificity when used in combination with other RF or CCP antibody assays
15384 ^b	Rheumatoid Factor Screen with Reflex to Titer, Synovial Fluid	Diagnose RA and determine prognosis
Sarcoidosis		
8561	Absolute Lymphocyte Count	Support diagnosis of sarcoidosis
683	Angiotensin Converting Enzyme	Support diagnosis of sarcoidosis
303	Calcium	Support diagnosis of sarcoidosis
1635	Calcium, 24-Hour Urine with Creatinine	Support diagnosis of sarcoidosis
6399	CBC (Includes Differential and Platelets)	Support diagnosis of sarcoidosis
375	Creatinine	Support diagnosis of sarcoidosis and determine extent of organ involvement

(Continued)

Table. Laboratory Tests for Classification and Diagnosis of Autoimmune Rheumatic and Related Diseases^a (Continued)

Test Code	Test Name	Clinical Use
7083	Immunoglobulins Panel, Serum Includes IgA, IgG, and IgM.	Support diagnosis of sarcoidosis
294	Urea Nitrogen (BUN)	Support diagnosis of sarcoidosis and determine extent of organ involvement
Sjögren Syndrome		
90077 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Sjögren's Panel 1 Includes ANA screen (IFA) with reflex to titer and pattern; rheumatoid factor; and SS-A and SS-B antibodies.	Diagnose Sjögren syndrome
19880 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Sjögren's Panel 2 Includes ANA screen (IFA) with reflex to titer and pattern (test code 249); mitochondrial antibody screen with reflex to titer (test code 259); rheumatoid factor (test code 4418); and SS-A (test code 38568), SS-B (test code 38569), and thyroid peroxidase (test code 5081) antibodies.	Diagnose Sjögren syndrome
4418	Rheumatoid Factor	Diagnose Sjögren syndrome
38568	Sjögren's Antibody (SS-A)	Diagnose Sjögren syndrome
38569	Sjögren's Antibody (SS-B)	Diagnose Sjögren syndrome
7832	Sjögren's Antibodies (SS-A, SS-B)	Diagnose Sjögren syndrome
Spondyloarthropathies		
4420	C-Reactive Protein (CRP)	Diagnose axial spondyloarthritis and assess disease activity
528	HLA-B27 Antigen	Diagnose SpA
15584	HLA-B27 DNA Typing	Diagnose SpA
Systemic Lupus Erythematosus		
8561	Absolute Lymphocyte Count	Detect lymphopenia
90072 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Lupus Panel 1 Includes ANA screen (IFA) with reflex to titer and pattern and chromatin (nucleosomal), dsDNA, and Sm antibodies.	Diagnose SLE
29839 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Lupus Panel 2 Includes ANA screen (IFA) with reflex to titer and pattern; also includes dsDNA, scleroderma (Scl-70), Sm, Sm/RNP, SS-A, and SS-B antibodies.	Diagnose SLE
19881 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern/ Lupus Panel 3 Includes ANA screen (IFA) with reflex to titer and pattern; also includes chromatin (nucleosomal), dsDNA, RNP, Sm, SS-A, and SS-B antibodies; complement components C3 and C4 and total complement (CH50).	Diagnose SLE
10716	ANA Screen, IFA, with Reflex to Titer and Pattern/ Lupus Panel 4 Includes ANA screen (IFA) with reflex to titer and pattern; dsDNA, rheumatoid factor, ribosomal P, Scl-70, Sm, Sm/RNP, SS-A, SS-B, and thyroid peroxidase antibodies; and complement components C3 and C4.	Diagnose SLE

(Continued)

Table. Laboratory Tests for Classification and Diagnosis of Autoimmune Rheumatic and Related Diseases^a (Continued)

Test Code	Test Name	Clinical Use
37491 ^{b,c}	ANA Screen, IFA, with Reflex to Titer and Pattern/ Lupus Panel 5 Includes ANA screen (IFA) with reflex to titer and pattern (test code 249); also includes actin (IgG, test code 15043), gastric parietal cell (test code 15114), rheumatoid factor (test code 4418), ribosomal P (test code 34283), Scl-70 (test code 4942), Sm (test code 37923), Sm/RNP (test code 38567), SS-A (test code 38568), SS-B (test code 38569), and thyroid peroxidase (test code 5081) antibodies; dsDNA (Crithidia, test code 37092); mitochondrial (test code 259), myocardial (test code 261), reticulin (test code 37520), and striated muscle antibody (test code 266) screens with reflex to titers; and C3 and C4 complement components (test code 5704).	Diagnose SLE
30340	Beta-2-Glycoprotein I Antibodies (IgG, IgA, IgM)	Diagnose SLE
7352	Cardiolipin Antibodies (IgA, IgG, IgM)	Diagnose SLE
34088	Chromatin (Nucleosomal) Antibody	Diagnose SLE
37859	Complement Component C3, C4, CH50	Diagnose SLE
361	Direct Antiglobulin Test (DAT)	Determine presence of autoimmune hemolytic anemia
255	DNA (ds) Antibody	Diagnose SLE
427	Erythropoietin	Determine presence of hemolytic anemia
19654	Lupus Anticoagulant and Antiphospholipid Confirmation (non-Coumadin) with Consultation	Diagnose SLE
17725(X)	Lupus Activity Panel 2 Includes complement component C3 and C4 and high avidity dsDNA antibody.	Diagnose SLE
91740	Platelet Antibody, Direct (IgG)	Detect autoimmune thrombocytopenia
34283	Ribosomal P Antibody	Diagnose neuropsychiatric SLE
19887	RNP Antibody	Diagnose SLE or MCTD
38567	Sm/RNP Antibody	Diagnose SLE or MCTD
37923	Sm Antibody	Diagnose SLE
937	White Blood Cell Count (WBC)	Determine presence of leukopenia
Systemic Sclerosis		
90073 ^b	ANA Screen, IFA with Reflex to Titer and Pattern/ Systemic Sclerosis Panel 1 Includes ANA screen (IFA) with reflexes to titer and pattern and centromere B and Scl-70 antibodies.	Diagnose systemic sclerosis
94646 ^d	Anti-PM/Scl-100 Antibody, EIA	Determine prognosis
16088	Centromere B Antibody	Diagnose limited cutaneous systemic sclerosis (CREST)
18855	Ku Autoantibodies	Predict muscle and joint involvement and digital vasculopathy-related complications
19899	RNA Polymerase III Antibody	Diagnose systemic sclerosis
4942	Scleroderma Antibody (Scl-70)	Diagnose systemic sclerosis

(Continued)

Table. Laboratory Tests for Classification and Diagnosis of Autoimmune Rheumatic and Related Diseases^a (Continued)

Test Code	Test Name	Clinical Use
94685 ^c	Systemic Sclerosis 12 Antibodies Panel 2 Includes centromere protein (CENP)-A, CENP-B, PM-Scl100, PM-Scl75, RP11 (RNA polymerase III), RP155 (RNA polymerase III), Scl-70, Th/To, U1 snRNP RNP70k, U1 snRNP RNP A, U1 snRNP RNP C, and U3-snRNP (fibrillarin) antibodies.	Diagnose systemic sclerosis
Systemic Vasculitis		
10547 ^b	ANA Multiplex with Reflex to dsDNA	Differentially diagnose ARDs
70159 ^b	ANCA Screen with MPO and PR3, with Reflex to ANCA Titer	Differentiate types of systemic vasculitis
4420	C-Reactive Protein (CRP)	Identify inflammatory conditions
375	Creatinine	Assess renal function
425	Eosinophil Count, Blood	Assess eosinophilia
427	Erythropoietin	Determine presence of hemolytic anemia
809	Sed Rate by Modified Westergren	Determine disease severity
8796	Myeloperoxidase Antibody (MPO)	Diagnose ANCA-associated vasculitides
723	Platelet Count, EDTA	Assess thrombocytopenia
34151	Proteinase-3 Antibody	Diagnose ANCA-associated vasculitides
294	Urea Nitrogen (BUN)	Assess renal function
8563	Urinalysis, Microscopic	Determine presence of microhematuria
937	White Blood Cell Count (WBC)	Assess leukopenia
ANA Screening Panels		
249 ^b	ANA Screen, IFA, with Reflex to Titer and Pattern	Differentially diagnose ARDs
16814 ^b	ANA Screen, IFA, with Reflex Titer and Pattern, and Reflex to Multiplex 11 Ab Cascade Antibody cascade includes chromatin, dsDNA, RNP, Sm, Sm/RNP antibodies; if all 5 antibodies are negative, reflex to SS-A, SS-B, Scl-70, and Jo-1 antibodies; if all 4 of these antibodies are negative, reflex to ribosomal P and centromere B antibodies.	Differentially diagnose ARDs
19946 ^b	ANA Multiplex with Reflex to 11 Antibody Cascade Includes ANA multiplex test with reflex to chromatin, dsDNA, RNP, Sm, Sm/RNP antibodies; if all 5 antibodies are negative, reflex to SS-A, SS-B, Scl-70, and Jo-1 antibodies; if all 4 of these antibodies are negative, reflex to ribosomal P and centromere B antibodies.	Differentially diagnose ARDs
94954 ^b	ANA Screen, IFA, Reflex Titer/Pattern, Reflex Mplx 11 Ab Cascade with IdentRA [®]	Differentially diagnose ARDs

^a Panel components may be ordered separately.

^b Reflex tests are performed at an additional charge and are associated with an additional CPT code.

^c This test was developed and its analytical performance characteristics have been determined by Quest Diagnostics. It has not been cleared or approved by the US Food and Drug Administration. This assay has been validated pursuant to the CLIA regulations and are used for clinical purposes.

^d This test was developed and its performance characteristics validated by Rheumatology Diagnostic Laboratory. There is no FDA approved assay for this test. As a lab developed test (LDT), approval or clearance by the FDA is not required. This test may be used for clinical purposes and should not be regarded as investigational or for research.

References

- Neogi T, Jansen TL, Dalbeth N, et al. 2015 Gout classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheumatol.* 2015;67:2557-2568.
- Zhang W, Doherty M, Bardin T, et al. European League Against Rheumatism recommendations for calcium pyrophosphate deposition. part I: terminology and diagnosis. *Ann Rheum Dis.* 2011;70:563-570.

3. Terkeltaub R. Calcium crystal disease: calcium pyrophosphate dihydrate and basic calcium phosphate. In: Firestein GS, Budd RC, Gabriel SE, et al, eds. *Kelley's Textbook of Rheumatology*. 9th ed. Philadelphia, PA: Elsevier Saunders; 2013:1577-1596.
4. Petty RE, Southwood TR, Manners P, et al. International League of Associations for Rheumatology classification of juvenile idiopathic arthritis: second revision, Edmonton, 2001. *J Rheumatol*. 2004;31:390-392.
5. Ortega-Hernandez OD, Shoenfeld Y. Mixed connective tissue disease: an overview of clinical manifestations, diagnosis and treatment. *Best Pract Res Clin Rheumatol*. 2012;26:61-72.
6. Cappelli S, Bellando Randone S, Martinović D, et al. "To be or not to be," ten years after: evidence for mixed connective tissue disease as a distinct entity. *Semin Arthritis Rheum*. 2012;41:589-598.
7. Bennett R. Overlap syndromes. In: Firestein GS, Budd RC, Gabriel SE, et al., eds. *Kelley's Textbook of Rheumatology*. 9th ed. Philadelphia, PA: Elsevier Saunders; 2013:1431-1451.
8. Gunnarsson R, El-Hage F, Aaløkken TM, et al. Associations between anti-Ro52 antibodies and lung fibrosis in mixed connective tissue disease. *Rheumatology (Oxford)*. 2016;55:103-108.
9. Lundberg IE, Tjärnlund A, Bottai M, et al. 2017 European League Against Rheumatism/American College of Rheumatology classification criteria for adult and juvenile idiopathic inflammatory myopathies and their major subgroups. *Ann Rheum Dis*. 2017;76:1955-1964.
10. Lega JC, Fabien N, Reynaud Q, et al. The clinical phenotype associated with myositis-specific and associated autoantibodies: a meta-analysis revisiting the so-called antisynthetase syndrome. *Autoimmun Rev*. 2014;13:883-891.
11. Hengstman GJ, van Brenk L, Vree Egberts WT, et al. High specificity of myositis specific autoantibodies for myositis compared with other neuromuscular disorders. *J Neurol*. 2005;252:534-537.
12. Vázquez-Abad D, Rothfield NF. Sensitivity and specificity of anti-Jo-1 antibodies in autoimmune diseases with myositis. *Arthritis Rheum*. 1996;39:292-296.
13. Aggarwal R, Oddis CV, Goudeau D, et al. Anti-signal recognition particle autoantibody ELISA validation and clinical associations. *Rheumatology (Oxford)*. 2015;54:1194-1199.
14. Nakashima R, Imura Y, Hosono Y, et al. The multicenter study of a new assay for simultaneous detection of multiple anti-aminoacyl-tRNA synthetases in myositis and interstitial pneumonia. *PLoS One*. 2014;9:e85062. doi:10.1371/journal.pone.0085062
15. Love LA, Leff RL, Fraser DD, et al. A new approach to the classification of idiopathic inflammatory myopathy: myositis-specific autoantibodies define useful homogeneous patient groups. *Medicine (Baltimore)*. 1991;70:360-374.
16. Aletaha D, Neogi T, Silman AJ, et al. 2010 rheumatoid arthritis classification criteria: an American College of Rheumatology/ European League Against Rheumatism collaborative initiative. *Arthritis Rheum*. 2010;62:2569-2581.
17. Maksymowych WP, Nades SJ, Bykerk V, et al. Serum 14-3-3 η is a novel marker that complements current serological measurements to enhance detection of patients with rheumatoid arthritis. *J Rheumatol*. 2014;41:2104-2113.
18. Dubucquoi S, Solau-Gervais E, Lefranc D, et al. Evaluation of anti-citrullinated filaggrin antibodies as hallmarks for the diagnosis of rheumatic diseases. *Ann Rheum Dis*. 2004;63:415-419.
19. Greiner A, Plischke H, Kellner H, et al. Association of anti-cyclic citrullinated peptide antibodies, anti-citrullin antibodies, and IgM and IgA rheumatoid factors with serological parameters of disease activity in rheumatoid arthritis. *Ann NY Acad Sci*. 2005;1050:295-303.
20. Sauerland U, Becker H, Seidel M, et al. Clinical utility of the anti-CCP assay: experiences with 700 patients. *Ann NY Acad Sci*. 2005;1050:314-318.
21. Suzuki K, Sawada T, Murakami A, et al. High diagnostic performance of ELISA detection of antibodies to citrullinated antigens in rheumatoid arthritis. *Scand J Rheumatol*. 2003;32:197-204.
22. Bizzaro N, Mazzanti G, Tonutti E, et al. Diagnostic accuracy of the anti-citrulline antibody assay for rheumatoid arthritis. *Clin Chem*. 2001;47:1089-1093.
23. Vallbracht I, Rieber J, Oppermann M, et al. Diagnostic and clinical value of anti-cyclic citrullinated peptide antibodies compared with rheumatoid factor isotypes in rheumatoid arthritis. *Ann Rheum Dis*. 2004;63:1079-1084.
24. Lee DM, Schur PH. Clinical utility of the anti-CCP assay in patients with rheumatic diseases. *Ann Rheum Dis*. 2003;62:870-874.
25. Forslind K, Ahlmén M, Eberhardt K, et al. Prediction of radiological outcome in early rheumatoid arthritis in clinical practice: role of antibodies to citrullinated peptides (anti-CCP). *Ann Rheum Dis*. 2004;63:1090-1095.
26. Wener MH, Hutchinson K, Morishima C, et al. Absence of antibodies to cyclic citrullinated peptide in sera of patients with hepatitis C virus infection and cryoglobulinemia. *Arthritis Rheum*. 2004;50:2305-2308.
27. Jansen AL, van der Horst-Bruinsma I, van Schaardenburg D, et al. Rheumatoid factor and antibodies to cyclic citrullinated peptide differentiate rheumatoid arthritis from undifferentiated polyarthritis in patients with early arthritis. *J Rheumatol*. 2002;29:2074-2076.
28. Maksymowych WP, van der Heijde D, Allaart CF, et al. 14-3-3 η is a novel mediator associated with the pathogenesis of rheumatoid arthritis and joint damage. *Arthritis Res Ther*. 2014;16:R99.
29. Kilani RT, Maksymowych WP, Aitken A, et al. Detection of high levels of 2 specific isoforms of 14-3-3 proteins in synovial fluid from patients with joint inflammation. *J Rheumatol*. 2007;34:1650-1657.
30. Wolfe F. Comparative usefulness of C-reactive protein and erythrocyte sedimentation rate in patients with rheumatoid arthritis. *J Rheumatol*. 1997;24:1477-1485.
31. American Thoracic Society. Statement on sarcoidosis: joint statement of the American Thoracic Society (ATS), the European Respiratory Society (ERS) and the World Association of Sarcoidosis and Other Granulomatous Disorders (WASOG) adopted by the ATS Board of Directors and by the ERS Executive Committee, February 1999. *Am J Respir Crit Care Med*. 1999;160:736-755.
32. Heinle R, Chang C. Diagnostic criteria for sarcoidosis. *Autoimmun Rev*. 2014;13:383-387.

33. Sweiss NJ, Baughman RP. Sarcoidosis. In: Firestein GS, Budd RC, Gabriel SE, et al., eds. *Kelley's Textbook of Rheumatology*. 9th ed. Philadelphia, PA: Elsevier Saunders; 2013:1898-1906.
34. Shiboski CH, Shiboski SC, Seror R, et al. 2016 American College of Rheumatology/European League Against Rheumatism classification criteria for primary Sjogren's syndrome: a consensus and data-driven methodology involving three international patient cohorts. *Ann Rheum Dis*. 2017;76:9-16.
35. Shiboski SC, Shiboski CH, Criswell L, et al. American College of Rheumatology classification criteria for Sjogren's syndrome: a data-driven, expert consensus approach in the Sjogren's International Collaborative Clinical Alliance cohort. *Arthritis Care Res (Hoboken)*. 2012;64:475-487.
36. Seror R, Gottenberg JE, Devauchelle-Pensec V, et al. European League Against Rheumatism Sjogren's Syndrome Disease Activity Index and European League Against Rheumatism Sjogren's Syndrome Patient-Reported Index: a complete picture of primary Sjogren's syndrome patients. *Arthritis Care Res (Hoboken)*. 2013;65:1358-1364.
37. Rudwaleit M, van der Heijde D, Landewé R, et al. The Assessment of SpondyloArthritis International Society classification criteria for peripheral spondyloarthritis and for spondyloarthritis in general. *Ann Rheum Dis*. 2011;70:25-31.
38. Rudwaleit M, van der Heijde D, Landewé R, et al. The development of Assessment of SpondyloArthritis international Society classification criteria for axial spondyloarthritis (part II): validation and final selection. *Ann Rheum Dis*. 2009;68:777-783.
39. Petri M, Orbai AM, Alarcón GS, et al. Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum*. 2012;64:2677-2686.
40. Dall'era M, Wofsy D. Clinical features of systemic lupus erythematosus. In: Firestein GS, Budd RC, Gabriel SE, et al, eds. *Kelley's Textbook of Rheumatology*. 9th ed. Philadelphia, PA: Elsevier Saunders; 2013:1283-1303.
41. Braun J, Bollow M, Remlinger G, et al. Prevalence of spondylarthropathies in HLA-B27 positive and negative blood donors. *Arthritis Rheum*. 1998;41:58-67.
42. Cervera R, Viñas O, Ramos-Casals M, et al. Anti-chromatin antibodies in systemic lupus erythematosus: a useful marker for lupus nephropathy. *Ann Rheum Dis*. 2003;62:431-434.
43. Kavanaugh A, Tomar R, Reveille J, et al. Guidelines for clinical use of the antinuclear antibody test and tests for specific autoantibodies to nuclear antigens. *Arch Pathol Lab Med*. 2000;124:71-81.
44. Stinton LM, Fritzler MJ. A clinical approach to autoantibody testing in systemic autoimmune rheumatic disorders. *Autoimmun Rev*. 2007;7:77-84.
45. Ho RC, Thiaghu C, Ong H, et al. A meta-analysis of serum and cerebrospinal fluid autoantibodies in neuropsychiatric systemic lupus erythematosus [published online ahead of print October 23, 2015]. *Autoimmun Rev*. 2016;15:124-138. doi:10.1016/j.autrev.2015.10.003
46. van den Hoogen F, Khanna D, Fransen J, et al. 2013 classification criteria for systemic sclerosis: an American College of Rheumatology/European League against Rheumatism collaborative initiative. *Arthritis Rheum*. 2013;65:2737-2747.
47. Solomon DH, Kavanaugh AJ, Schur PH, et al. Evidence-based guidelines for the use of immunologic tests: antinuclear antibody testing. *Arthritis Rheum*. 2002;47:434-444.
48. Bunn CC, Denton CP, Shi-Wen X, et al. Anti-RNA polymerases and other autoantibody specificities in systemic sclerosis. *Br J Rheumatol*. 1998;37:15-20.
49. Spencer-Green G, Alter D, Welch HG. Test performance in systemic sclerosis: anti-centromere and anti-Scl-70 antibodies. *Am J Med*. 1997;103:242-248.
50. Leavitt RY, Fauci AS, Bloch DA, et al. The American College of Rheumatology 1990 criteria for the classification of Wegener's granulomatosis. *Arthritis Rheum*. 1990;33:1101-1107.
51. Masi AT, Hunder GG, Lie JT, et al. The American College of Rheumatology 1990 criteria for the classification of Churg-Strauss syndrome (allergic granulomatosis and angiitis). *Arthritis Rheum*. 1990;33:1094-1100.
52. Savage COS, Harper L. Antineutrophil cytoplasm antibody-associated vasculitis. In: Firestein GS, Budd RC, Gabriel SE, et al, eds. *Kelley's Textbook of Rheumatology*. 9th ed. Philadelphia, PA: Elsevier Saunders; 2013:1481-1497.
53. Jennette JC, Falk RJ, Bacon PA, et al. 2012 revised International Chapel Hill Consensus Conference nomenclature of vasculitides. *Arthritis Rheum*. 2013;65:1-11.
54. Savige J, Dimech W, Fritzler M, et al. Addendum to the International Consensus Statement on testing and reporting of antineutrophil cytoplasmic antibodies: quality control guidelines, comments, and recommendations for testing in other autoimmune diseases. *Am J Clin Pathol*. 2003;120:312-318.
55. Papp M, Altorjay I, Lakos G, et al. Evaluation of the combined application of ethanol-fixed and formaldehyde-fixed neutrophil substrates for identifying atypical perinuclear antineutrophil cytoplasmic antibodies in inflammatory bowel disease. *Clin Vaccine Immunol*. 2009;16:464-470.
56. Comarmond C, Pagnoux C, Khellaf M, et al. Eosinophilic granulomatosis with polyangiitis (Churg-Strauss): clinical characteristics and long-term followup of the 383 patients enrolled in the French Vasculitis Study Group cohort. *Arthritis Rheum*. 2013;65:270-281.
57. Reese GE, Constantinides VA, Simillis C, et al. Diagnostic precision of anti-Saccharomyces cerevisiae antibodies and perinuclear antineutrophil cytoplasmic antibodies in inflammatory bowel disease. *Am J Gastroenterol*. 2006;101:2410-2422.
58. Schönemarck U, Lamprecht P, Csernok E, et al. Prevalence and spectrum of rheumatic diseases associated with proteinase 3-antineutrophil cytoplasmic antibodies (ANCA) and myeloperoxidase-ANCA. *Rheumatology (Oxford)*. 2001;40:178-184.
59. Bossuyt X, Cohen Tervaert JW, Arimura Y, et al. Position paper: revised 2017 international consensus on testing of ANCAs in granulomatosis with polyangiitis and microscopic polyangiitis. *Nat Rev Rheumatol*. 2017;13:683-692.
60. Kaldas A, Warraich I, Prabhakar SS. ANCA associated glomerulonephritis-an in-depth review. *J Nephrol Ther*. 2013;4. doi:10.4172/2161-0959.1000147
61. Wu YY, Hsu TC, Chen TY, et al. Proteinase 3 and dihydroliipoamide dehydrogenase (E3) are major autoantigens in hepatitis C virus (HCV) infection. *Clin Exp Immunol*. 2002;128:347-352.

